

**5 min TA/Blunt-Zero
Cloning Kit**

C601



Instruction for Use
Version 21.1

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01/Product Description

5 min TA/Blunt-Zero Cloning Kit is a second generation TOPO cloning kit that contains a second generation Topoisomerase, a vector containing the suicide gene *ccdB* and a blunt end factor. Combining with the optimal buffer, the second generation of Topoisomerase provides a highly efficient, 5 min, one-step cloning strategy at room temperature. This product using a vector containing the suicide gene *ccdB*, when the insert is successfully ligated to the vector, the correct expression of *ccdB* is destroyed, and the host cell can grow normally, otherwise the host cell cannot grow normally, thereby achieving “zero” background. Containing a blunt end factor, 5 min TA/Blunt-Zero Cloning Kit is compatible with both TA clones and blunt clones.

02/Components

Components	C601-01 (25 rxns)	C601-02 (50 rxns)
5 × TA/Blunt-Zero Cloning Mix ^a	25 µl	2 × 25 µl
500 bp Control insert (20 ng/µl)	5 µl	10 µl
M13 Primer Mix (10 µM) ^b	200 µl	400 µl

a. Contains Topoisomerase and pCE2 TA/Blunt-zero Vector (double resistance: Amp^r, Kan^r)

b. Contains M13 Forward Primer and M13 Reverse Primer

03/Storage

Store at -30 ~ -15°C, and transport at ≤0°C.

04/Applications

It is suitable for ligation of blunt ended and A-Tailed PCR products.

05/Notes

For research use only. Not for use in diagnostic procedures.

06/Experiment Process

1. Summary of the Experimental Process

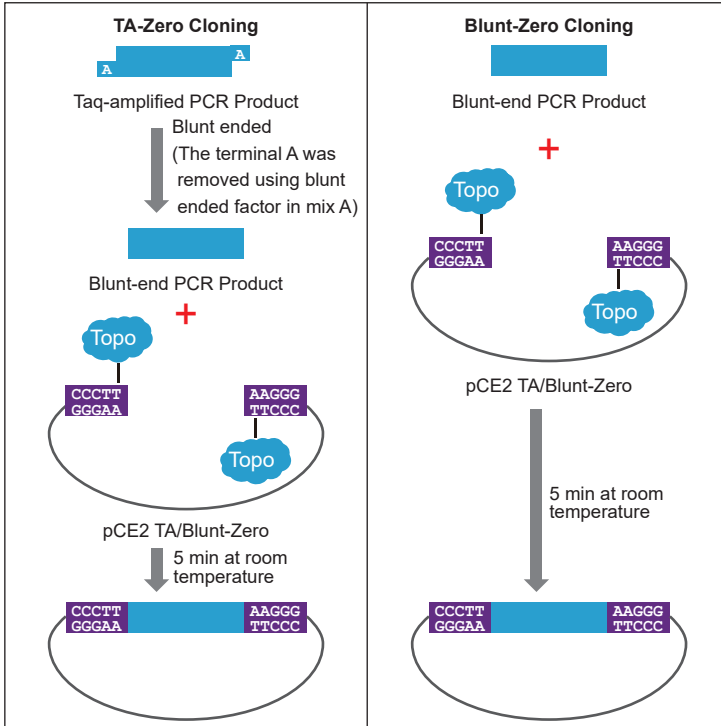


Figure A

Figure B

Process Summary of 5 min TA/Blunt-Zero Cloning Kit

Fig A: TA-Zero Cloning

- Add the amplification product which 3' end containing A of Taq (Vazyme #P211) to 5 × TA/Blunt-Zero Cloning Mix, incubate at room temperature for 5 min.
- The blunt-end factor in Mix removes the A-base at the end of the amplification product to form a blunt-ended product.
- 5'-OH of the blunt-end product attacks the phosphate bond between the TOPO enzyme and the vector, the TOPO enzyme is released, and the vector forms a circular recombinant with the blunt-ended product.

Fig B: Blunt-Zero Cloning

- Amplification products (blunt ends) of high-fidelity enzymes (Vazyme #P505) were added to 5 × TA/Blunt-Zero Cloning Mix and incubated at room temperature for 5 min.
- 5'-OH of the blunt-end product attacks the phosphate bond between the TOPO enzyme and the vector, the TOPO enzyme is released, and the vector forms a circular recombinant with the blunt-ended product.

2. PCR Product Preparation

- a. Primer requirements: the 5' end of the primer cannot be phosphorylated.
- b. Enzyme selection: It is recommended to use Taq Master Mix (Vazyme #P211) or Phanta series products (Vazyme #P505).
- c. Product requirements: Please ensure the integrity of the PCR amplification products; after the end of the amplification, the yield and quality of the product are detected by electrophoresis, if the product has only the target band, no non-specific band and primer dimers, it can be used directly, otherwise it is recommended to carry out gel recovery and purification. If the amplification template is plasmid, purification is recommended.

3. Ligation Reaction

Prepare the reaction mix:

Components	Volume
5 × TA/Blunt-Zero Cloning Mix	1 μl
Purified PCR Product	1 - 4 μl
ddH ₂ O	To 5 μl

Mix the bottom of the flick tube, collect all the liquid at the bottom of the centrifuge tube at low speed and react at room temperature (20 ~ 37°C) for 5 min. After the reaction was over, the tube was placed on ice.

Recommended reaction conditions:

a. The optimum amount of inserts used = $[0.05 \times \text{fragment base pairs}] \text{ ng}$;

For example, when the insert is 1,000 bp, the optimum amount is $[0.05 \times 1,000] \text{ ng}$, that is, 50 ng. Due to the wide range of compatibility of the inserts of this product, you can also use the recommended dosages in the table below:

Inserts Size	Recommended Dosages
0.5 - 1 kb	5 - 60 ng
1 - 2 kb	60 - 110 ng
2 - 5 kb	110 - 260 ng
>5 kb	>260 ng

▲ Nanodrop, Onedrop, etc. are recommended for concentration determination.

b. Reaction Temperature: This product has high compatibility with reaction temperature, so the reaction can be performed at room temperature (20 ~ 37°C). A PCR instrument controlled temperature of 25°C is recommended.

c. Reaction Time: React for 5 min.

4. Transformation

This product is compatible with many conventional competent cells, such as DH5 α competent cell (Vazyme #C502); Fast-T1 competent cell (Vazyme #C505).

▲ It is recommended to use Fast-T1 competent cell (Vazyme #C505) for subsequent transformation experiments. The cells are the fastest growing competent cells (clones can be seen 8h after plating), and the transformation efficiency is high, saving screening time.

5. Positive Clone Identification

a. PCR identification of the bacterial colony and solution: pick a single colony to 10 μ l of ddH $_2$ O as a template; mix well as template; Recommended use of 2 \times Rapid Taq Master Mix (Vazyme #P222).

Reaction System:

Components	Volume
2 \times Taq Master Mix	10 μ l
M13 Primer Mix	2 μ l
Bacterial Solution	2 μ l
ddH $_2$ O	to 20 μ l

Reaction Procedure:

Temperature	Time	Cycles
95°C	3 min	} 35 cycles
95°C	15 sec	
55°C	15 sec	
72°C	15 sec/kb	
72°C	5 min	

b. Enzyme Digestion Analysis: According to the experimental design, select the appropriate restriction endonuclease to identify

c. Identification of Plasmid Size: Picking a single clone, after plasmid extraction, electrophoresis observation of plasmid size identification

d. Sequencing Analysis: Directly pick the monoclonal sequencing identification, sequencing primers can choose M13 Forward Primer, M13 Reverse Primer or Designed by yourself.

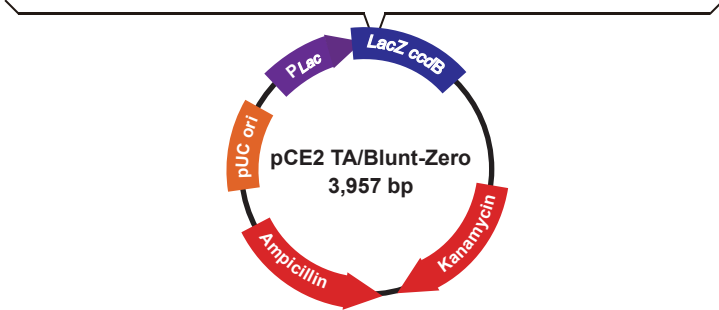
07/Attachment: Sequence Information of Vector

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      M13 Reverse Primer
325  CACAGGAAAC AGCTATGACC ATGATTACGC CAAGCTCAGA ATTAACCCCTC ACTAAAGGTA
      GTGTCCTTTG TCGATACTGG TACTAATGCG GTTCGAGTCT TAATTGGGAG TGATTTCAT

      EcoRI TOPO TOPO EcoRI
385  CTAGTCCTGC AGGTTTAAAC GAATTCGCC TT AAGGGCGA ATTCGCGGCC
      GATCAGGACG TCCA AATTTG CTTAAGCGGG AA PCR Product TTCCC GCT TAAGCGCCGG

      M13 Forward Primer
435  GCTAAATTC A ATTCCGCCCTA TAGTGAATCG TATTACAATT CACTGGCC GTCGTTTTACAA
      CGATTTAAGT TAAGCGGGAT ATCACTTAGC ATAATGTTAA GTGACCGG CAGCAAATGTT
  
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Lac promoter: bases 217 - 338

LacZ ccdB fragment: bases 339 - 932

M13 Reverse primer site: bases 327 - 343

TOPO binding site (left): bases 412 - 416

TOPO binding site (right): bases 417 - 421

M13 Forward primer site: bases 476 - 492

Kanamycin resistance ORF: bases 1,281 - 2,075

Ampicillin resistance ORF (C): bases 2,226 - 3,239

pUC origin: bases 3,284 - 3,957

(C): complementary strand

For more information about pCE2 TA/Blunt-Zero Vector, Please refer to www.vazyme.com



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